DNA extraction from Tissue (mouse tail)

Preparation of test samples
Cut 3～5mm of the mouse tail (c.a.10mg) and put it in the micro test tube
(usually use 0.2ml or 0.5ml tubes for PCR)

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Reagents
Mix the CellEase A, B and distilled water
(20 µl CellEase A, 20 µl CellEase B, 60 µl distilled water)

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Add 100 µl of the mixture to the samples.

↓

Incubate at 72℃ for 6 minutes
Then incubate at 94℃ for 3 minutes

↓

Transfer 6 µl of extracts to PCR reaction mixture
and amplify the target DNA fragment

PCR

6.0 µl	Test sample
5.0 µl	×10 buffer(+Mg^{2+})
5.0 µl	dNTPs
1.0 µl	Forward Primer (10pmol/µl)
1.0 µl	Reverse Primer (10pmol/µl)
0.5 µl	Ex Taq (5 U/µl)

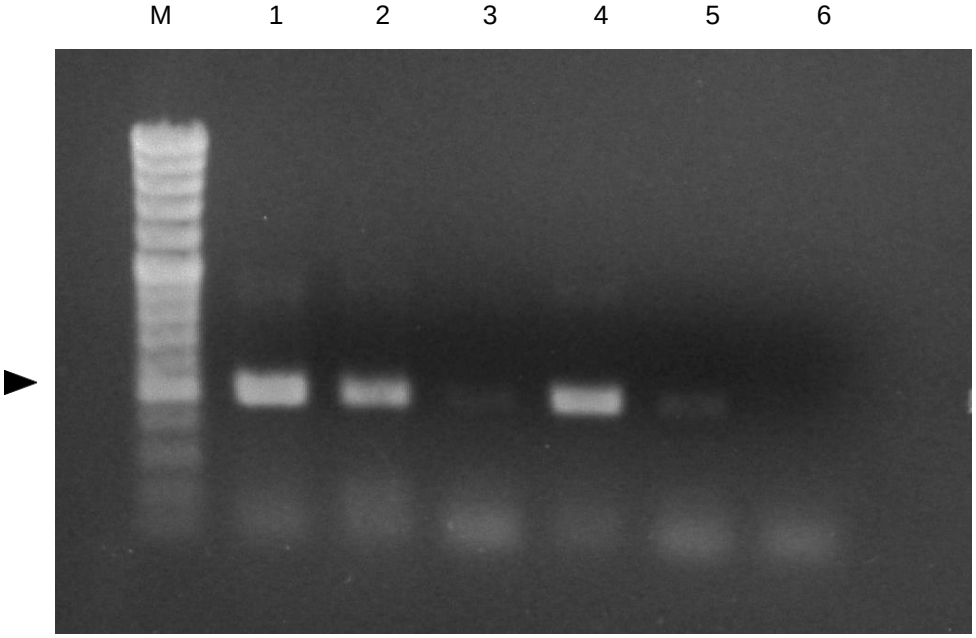
Fill up to 50 µl by distilled water

PCR Cycle

94℃	1min	
94℃	30sec	35 Cycles
55℃	30sec	
72℃	60sec	
72℃	4min	

Primers: Mouse β-globin gene (494bp)

＜Comparison with conventional CellEase kit ＞



M	Marker (100bp ladder)	
1	DNA extract by using CellEase Tissue Ⅱ	conc.
2	DNA extract by using CellEase Tissue Ⅱ	×10
3	DNA extract by using CellEase Tissue Ⅱ	×100
4	DNA extract by using conventional CellEase	conc.
5	DNA extract by using conventional CellEase	×10
6	DNA extract by using conventional CellEase	×100

※ The protocol of conventional CellEase and CellEase II kit were followed by the instruction manual respectively.

※ The thickness of the DNA bands were depending upon the amount of test samples and a parts of tissue including bone, skin or fat.

The clear DNA bands were detected from more than ×10 dilution of DNA extracts by using CellEase Tissue Ⅱ

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